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THE PREVALENCE OF *Candida albican* IN GARRI SOLD IN OZORO

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ABSTRACT

Garri is a fermented cassava product consumed as a staple food in most parts of West African. The prevalence of *Candidaalbican* in garri sold in ozoro was carried out to characterize predominant fungi associated with garri. Ten (10) different samples of garri were periodically bought from Ozoro central markets. The samples were labeled sample A-J. A total of three (3) fungi species were isolated from the test samples: *Aspergillus spp., Candida albican, and Mould.* The total heterotrophic plate count ranges from 2.4x10³cfu/ml to3.2x10³cfu/ml. *Candidaalbican* have the highest percentage occurrence of 62.5%, while Mould species have the least percentage occurrence of 12.5%. The pH of the samples ranged from 6.0 to 6.2. The occurrence of *Aspergillusspp*, mould and *Candidaalbican* indicates a strong likely hood of cross contamination between handling and utensils used during the garri fermentation or production processes of the exterior region.

INTRODUCTION

Nigeria is experiencing increase in population growth. The high population without equivalent increase in food production and availability to the citizens could result to malnutrition, disease outbreak and death. It is an important factor which will need attention by policy makers in Nigeria. Children and lactating mother needs and should be protein given adequate necessary nutrients or balance diet. According to (Jonathan et al., 2011), effect of poor food is

termed or known as malnutrition, low productivity level among mothers and children. Among factors affecting inadequate food suffered by citizens is low socioeconomic level of the people which is a serious problem. The above problems listed prompted campaign for increase production, utilization and consumption of traditional foods includes which fresh and processed cassava among the citizens, (FAO, 1998). Cassava mainly tubers consist carbohydrates (90% of dry weight basis) (Kay, 1973), 3% protein content and lack cysteine and methionine (Gomez, et al., 1985). The cassava leaves. however, are rich in protein; vitamin A and B are commonly consumed as a vegetable by Africans (Anonymous, and Kimen, et al., 2000). Cassava for garri production is harvested manually in the farm with the aid of a cutlass, flat iron steel (digger) and toe that usually inflicts various degrees of injuries on the root tubers of the cassava. After the cassava has been harvested. the root tubers are traveled to the market where they are heaped in 20s, 40s, 50s, 60s, 100s, and above for sales under humid and warm condition of the atmosphere. These practices predispose or exposed the root tubers to contamination and infestation by various groups of micro-organism (fungi) and it may expose public health hazards to citizens. Garri is consumed mainly as a main meal (Eba) or by taking it as a snack by soaking it in cold water with the addition of sugar or salt and groundnut, fried fish e.t.c. Garri is sometimes consumed with milk beverage. Various groups of microorganism such as bacteria, fungi e.t.c. have been reported to be associated with garri production and storage, also during distribution. Fungi can

grow and affect the nutritional and sensory properties or features of garri. Aflatoxins which is produced by fungi genera such as *Aspergillus* and *Penicllium*. Aflatoxins B₁, B₂, G₁, and G₂ are the most frequently encountered mycotoxins because they are produced by ubiquitous fungi genera.

Some fungi are pathogenic and can cause systemic illness e.g. Candida sp. Also, sporogenic fungi when ingested in the garri may come out sporulation to cause illness. In market, garri that is meant to be consumed by the public should be in a safe hand by the manufacturers. Fungi may be multicellular or unicellular (Banwart, 1974). Moulds belonging to the Eumycetes or True fungi whose growth on food is readily recognized by its fuzzy appearance. Yeasts on the other hand are unicellular fungi which are able to reproduce vegetatively by means of simple cells, which bud is commonly divided b fission (Pitt and Hocking, 1997). This study evaluates the prevalence of Candida albican on garri sold in Ozoro market.

Materials and Method Study Area

Ozoro is the headquarters of Isoko North Local Government Area of Delta State. It is one of the administrative units of the Isoko regions in Delta State Nigeria situated at Latitude: 5.5383 and Longitude: 6.2161 with approximate population of 13,411(at 2015) inhabitants and land mass of 1.136km². Ozoro falls within the southern tropical evergreen forest zone characterized by two climatic seasons. It comprises also of commercial activities and other municipal practices which causes environment pollution.

Collection of sample

10 Garri samples were bought from three different locations in Ozoro, Isoko North and Delta state. The samples were collected into sterile polythene and tied properly place of purchase and the samples were labeled A-J. and were transported to the laboratory, where analysis was carried out.

Materials

The material used in this study includes laboratory coat, gloves weighing balance, cotton wool foil paper measuring cylinder (50ml,250ml, 500ml), and beakers (50ml, 200ml,500ml), pipette, petri dishes test tubes

conical flask, Bunsen burner, potato dextrose Agar (PDA), wire loop, water etc.

Sterilization of Glasswares

The glasswares that were used for this project were washed with detergent, rinsed thoroughly and sterilized using autoclave at 121°c for 15 minutes.

Method Isolation of Fungi on Garri

The materials used in this study include laboratory coat, gloves weighing balance, cotton wool, slides, sterile universal container, foil paper. incubator. microscope, measuring cylinder (50ml. 250ml, 500ml). beakers (50ml, 200ml, 500ml) pipette, Petri dishes, test tubes, conical flask, Bunsen burner, potato dextrose Agar (PDA), wire loop and water.

Preparation of sterilewater

50 milliliters (50ml) of distilled water was pipette into a clean dry test tube plugged with cotton-wool and which was wrapped with aluminum foil. The test tubes were place into an autoclave and sterilized by autoclaving at 121°c for 15 minutes.

Preparation of media Potato Dextrose Agar (PDA)

Thirty-nine grams (39g) of PDA was weighted into a one (1) liter capacity of conical flask and was boiled and distributed into Mac Conkey bottles and autoclaved for 121°c for 15 minutes.

Sample Preparation

The samples (10g) brought from the market was suspended in a 50ml of sterile distilled water and was homogenized. The samples (A-J) were serially diluted under aseptic condition according to the method of Cheesbrough (2000). 0.1ml of 10², 10³, 10⁴, 10⁵, 10⁶ dilutions were transfer to plates of potato dextrose Agar (PDA).

Results and Discussion Results

The Fungi isolated from the garri samples are *Candida albicans*, *Aspergillus spp.* and *Moulds*.

Table 4.1: Shows the isolate, Cultural/morphological character

| CULTURAL/MORPHOLOGICAL CHARACTER | Isolates |
|--|------------------|
| Creamy white colour on SDA | Candida albicans |
| Whitish colour with fuzzy edges on SDA | Moulds |
| Black, white/yellow | Aspergillus spp. |

Table 4.2. Total Heterotrophic plate count

| Samples | CFU/ML | |
|---------|---------------------|--|
| Α | 3.2x10³ | |
| В | 2.4X10 ³ | |
| С | 3.6X10 ³ | |
| D | 3.3X10 ³ | |
| E | 2.6X10 ³ | |
| F | 2.9X10 ³ | |
| G | 2.7X10 ³ | |
| Н | 3.4X10 ³ | |
| 1 | 3.5X10 ³ | |
| J | 2.8X10 ³ | |

Table 4.3 pH of the samples

| Samples | рН | |
|---------|-----|--|
| Α | 6.0 | |
| В | 6.1 | |
| С | 6.1 | |
| D | 6.1 | |
| E | 6.1 | |
| F | 6.0 | |
| G | 6.2 | |
| Н | 6.1 | |
| I | 6.1 | |
| J | 6.1 | |
| | | |

Table 4.4 Frequency of occurrence of Fungi Isolates

| Isolates | No. | % Occurrence |
|--------------------------|---------|--------------|
| Aspergillius spp. | 4 | 25.0 |
| Candida albican Mould | 10 2 | 62.5 12.5 |
| Total | 16 | 100 |

DISCUSSION

These fungi are reportedly contaminants common agricultural commodities which includes, cereal (cassava tubers, soil and vegetation); Kunene, 1990. The pH of the samples in the present study were similar to the result obtained or achieved by Oyewole (2001) who reported pH to be 6.1. The lower count of Gram-negative fungi spore found in cooked garri and mineral storage at ambient temperature for up to 72hours in other to reduce the microbial population on garri (Holzapfel, 2002). Their

presence has been found to survive the high acidic conditions which develops towards the end fermentation; Oyewole, of (2001). This study highlighted that the high microbial number of garri as a source or result of spore forming, catalase positive. **Aspergillus** spp. However, deterioration in culture performance owning/due to one or more of these effects that adversely affects the fermentation process of garri production 2002). Mycotoxins, (Holzapfel, the toxic metabolites of certain fungi (moulds), may cause acute intoxication and longer term mutagenic, carcinogenic and tetratogenic effect when consumed largely.

Microbial counts of garri powder samples obtained from the three markets in Ozoro (Express market, small market and main market) ranges from 2.4x10³ – 3.6x10³cfu/ml, which are slightly lower than those found in powered garri samples analyzed Ofuya, and Akpobi, (1988).Ofuya (2001) reported that fuligo spp (white slime mould) appeared at the beginning of fufu fermentation and became extinct towards the end of the process. Predominant isolate identified from Express market sample were $3.2x10^{3}$ which reinforce result obtained by Okoli, (1979), Ejiofam and Okafor (1981), Ofuya, Akpoti, (1988).Amoa-Awua (1996). These microbial counts may have been derived from environments of the processing sites and the market. It is vital that garri should be produced and sold in an environment of high sanitary quality (Ofuya, and Akpoti, 1988). Fungi have been known to cause food borne illness when contaminated food is injected or consumed. (Gilbert and Perry, 1977; Hobbs and Gilbert, 1978 and Gilbert, 1979).

However, the ability of fungi to break down cassava tissue is likely to contribute to the detoxification of cassava during fermentation of garri by bringing about a greater contact between the cyanogenic glucosides and linamarase, which hydrolyses linamarin (Conn, 1969). Previous studies reported that the presence of Alcaligenes (genus of Gram-negative) in cassava fermentation is similar to the garri samples in this study. Oyewole, and Odunfa, 1990).

Relatively high numbers of yeast and moulds, which were not identified in this study, may constitute an additional quality and safetv risk in cassava fermentation (Kimayo, et al., 2000). Previous study by Westby and Twiddy (1992), reported the Isolation of fungi (Aspergillus spp., Mould and Candida albicans) in characterization of garri and fufu preparation procedure in Nigeria Samples similar to garri power samples analyzed in this study. This could be due to post processing problems, which include handling, air borne contaminants, storage conditions and packaging (Sanni, 1996). Inoculation with starter cultures does not provide an absolute quarantee against failure of fermentation processes; neither does it eliminate health

hazards associated with toxinogens, pathogens, toxic components and residence. LAB and yeast strains associated with fermented foods are capable of degrading anti-nutritional factors such as phylic and phenolic compounds, thus. when incorporated into food fermentation serve to upgrade the nutritional value (Holzapfel, 2002).

These fungi isolated could have got into garri during processing or as a result of being stored in contaminated storage facilities, reported presence of fungi in garri samples collected from various locations in Ozoro and presence of these organisms were attributed to poor handling techniques during processing, transport and storage. High moisture content in garri can be associated with the toasting method. In Ozoro, garri is toasted half way and then sun dried which may not be complete due to sudden change in weather that may lead to packing the garri from the sun. During sun drying, spores of fungi could deposited on the samples leading to germination when conditions are favorable for the organisms. These organisms are all storage fungi which proliferate when conditions are suitable, so garri sample with high moisture

content will lead to proliferation of these fungi.

CONCLUSION AND RECOMMENDATIONS Conclusion

The microbial average count of the garri analyzed ranges from 2.4x10³ to 3.6x10 ³cfu/ml. The significance indicates a strong like hood of cross contamination between handling and utensils used during the garri fermentation or production processes of the exterior region. The identification of randomly selected colonies from spore cont plates of the serial dilution resulted in up to 3.6x10³ of colonies identified as Aspergillus SPP, mould and Candida albicans.

RECOMMENDATIONS

It is hereby recommended that the identification of specific strains of pure starter culture for cassava fermentation needs to be obtained in order to determine if higher ethanol concentration would be produced. More work still need to be done or carryout on the production of cassava tubers into garri. And also an investigation on the use of mixed starter culture of ethanol to determine either higher ethanol concentrations would be produced.

REFERENCES

- Abe, M.O and Lindsay, R.C. (1978): Evidence for a Lactic Streptococcal Role in Nigeria Acid Cassava (Manihot esculenta crantz) Fermentation Journal of Food Protection 41, 781-784.
- Adeniji, M.O. (1976): Fungi Associated with the Deterioration of Garri Nigerian Journal of Plant Protection 2, 74-77.
- Al-Hassan, R.M. (1991): Cassava as a Food Security in Ghana Characterization of Village Level Processing and Marketing, in Root, Tuber Crops and Plantain Development in Ghana, Kumasi, Ghana, August 14-15, 1-118.
- Almazan, A.M. (1988): Selection of Cassava Varieties for Processing and Utilization. In: in Praise of Cassava. International Institute of Tropical Agriculture 1, 94-106.
- Amoa-Awua, W.K. and Jakobsen.M. (1995). The Role of Micro-Organism in the Fermentation of cassava. *Journal of Applied Microbiology* 79, 250-256.

- Amoa-Awua, W.K., Frisuad, J.C., Sefa-Dedehs.S., and Jakobsen.M. (1997). The Contribution of Moulds and Yeasts to the Fermentation of 'Agbelima' cassava dough.
- Anonymous, (1999a). Cassava puts Starch into Northern Province Review 3, 51-51.
- Anonymouse, (2000b).

 Championing the Cause of Cassava. United Nations:
 Food and Agriculture Organization., 1-4
- Anonymouse, (2000c). Cassava:
 A Mother Crop for Million.
 Indonesia.
 http://www.bath.ac.uk/Admin/Topics/t
- Best, R.(1988). Cassava Processing. The Latin American Experience: In: In praise of Cassava. International Institute of Tropical Agriculture 1, 37-52.
- Bokanga, M.199; Large Scale Ethanol Production from Cassava in Nigeria. Post Harvest System 1,1-6.
- Bawart, (1974). Multicellular or Unicellular Moulds

- Belonging to the Eumycetes Fungi. Yeast is Unicellular Fungi Produced Vegetationally by Means of Simple Cells 6, 30-83.
- Bokango, m.(1995); Biotechnology and Cassava Processing in Africa. Food Technology 49,86-90.
- Cannn, E.E.(1969). Cyanogenic Glucosides. *Journal of Agricultural and Food Chemistry 17,519-526.*
- Cheesbrough (2000): Serial Dilution of Garri Samples 56, 33-50.
- Dahniya, (1994): Cassava Production in selected Countries, Continent and the World 4, 5-7
- Datiniga, (1994): On average, farmers worldwide produce about 10 tons of Cassava Per hectares, but Yields can Reach 40 Tons per Hectares 24, 26-30.
- Ejofor, M.A.M and Okafor, N.(1981); Comparison of Pressed Cassava Pulp for Garri Making. 6,78-987.
- FAO.(1998). Mycotoxins Prevention and Control in Food Grain. Food and

- Agricultural Organization of the United Nations. Rome 3,456-456.
- Gilbert, R.J. and Parry. J.M. (1977). Serotypes of fungi from Outbreak of Food Poisoning and from Routie Foods. *Journal of Hygiene* 78.69-74.
- Grace, M.R (1977); Cassava processing. Rome: Food and Agriculture Organization, Plant Production and Protection series 3,1-1002.
- Grace, M.(1978); Cassava Processing Food and Agriculture Organization, 5-5.
- Hahn, S.K.(1989); An Overview of African Traditional Cassava Processing and Utilization. Outlook Agriculture 18, 110-118.
- Hocking, and pitt, (1997): Pathogenic Fungi; Cause Systemic Illness. Garri Meant to be Consumed by Public should be in safe Manufacturing state. Research Institute Technical Report 14, 60-127.

- Hobbs, B.C and Gilbert, Rj.(1978); Food Poisoning and Hygiene 4th ed. Edward Arnold, London, 25-26.
- Hunt, j.(1999). Cassava Puts Starch into Northern Province Economy, *Food Review*. 3,51-51.
- Halzapfel; (2002): Deterioration in Culture Performance Owing to Ane or more Effect which Affect garri Fermentation. International Institute of Tropical Agriculture 1,67-69.
- Jonathan. G., Ajayi. L. and Y. Omitade. (2011a). Nutritional Composition. Fungi and Aflatoxins Detection in Stored Garri Fermented, from South Western Nigerian. African Journal of Food Science 5(2): 105-110.
- Kay, D.E. (1977); Root crops. TPI Crop and Product Digest. Tropical Product Institute, London, 2.56-79.
- Kimaryo, V.M., Massawe, G.A. olasupa, N.A. and Holzapfel, W.H.(2000); The use of a starter culture in the fermentation of cassava for the production Ωf Kivunde: a traditional

- Tanzanian food product. *International Journal of Food Microbiology* 56,179-190.
- Kramer, J.M and Gilbert, R.J.(1989); Aflatoxins and other Fungi Species. In; Doyle M.P.eds, food Borne Pathogens, Marcel Dekker, New Yolk, 4, 21-70.
- Kunene, N.F. Geornaras, I., Von Holy A. and Hasting J.W.(2000).; Characterization and of Determination а Sorghum-based Fermented weaning food by Analysis of Soluble **Proteins** and Amplified Fragment Length Polymorphism Figer Printing. Applied and **Environmental** Microbiology 66. 1084-1092.
- Lancaster, P.A., Ingram, j.s., Lim, M.Y. and coursey, D.G.(1982). Traditional Cassava-based Foods: Survey of processing Techniques. Economic Botany 36:12-45.
- Madeley, J.(1993). Make Way for Supper Cassava. *Cereals* 949:2-6.

- Nestel, B.(1980). Cassava, A New Outlook for an Ancient crop. In Optima 1, 53-59.
- Nweka, F.I. Preliminary
 Observation from COSCA
 Phase I and II Survey.
 Tropical Root and Tuber
 Crops Bulletin 8:13-16.
- Nofal, j. (1999). Cassava; Ideal Filter Crop for Potato Farmers. In Farmers Weekly 12, 60-61.
- Ofuya, C.O. and Akpoto, P.(1988). Post-processing Microflora and the Shelf Stability of Garri Marketed in post Harcort. *Journal of Applied Bacteriology* 64: 389-394.
- Okafor, N. (1977).

 Microorganisms Associated with Cassava Fermentation for Garri Production.

 Journal of Applied Bacteriology 43: 279-284.
- Oyewole, O. (2001); Characteristics and Significance of Yeast Involvement in Cassava Fermentation of Fufu Production .International Journal of Food *Microbiology* 65: 213-218.
- Okoli, C.E. (1979). Microorganisms Associated

- with Cassava Fermentation for Garri Production Journal of Applied Bacteriology 42:279-284.
- Oyenira, J.O.(1979); Mould Development in Garri during Storage in Polyethylene and Hessian bags. Nigerian Stored Product Research Institute, Technical Report 11: 93-99
- Otto, J.A.(1998). African-wide Cassava Improvement Programme .In Praise of Cassava. International Institute of Tropical Agriculture 1: 67-69.
- Ofuya, C.O and Akpoti, O.(1988). Post Processing Microflora and Shelf Stability of Garri. *J. Applied Fungi.* 64, 389..-394.
- Oyewole, O. and Odunfa, SA.(1990). Characteristic and Distribution of Lactic Acid Bacteria in Cassava Fermentation during Fufu Production. *Journal of Applied Bacteriology* 68, 145-152.
- Sanni, M.O.(1996). Short Communication: The Stability of Stored Garri. International Journal of

Food Microbiology 29, 119-123.